

Your article proof EN-05-6161 from Environmental Entomology is available for download

=====

Dear Author,

This letter describes how to access and proof your article EN-05-6161 slated for an upcoming issue of Environmental Entomology. If you have any questions, contact the Environmental Entomology Issue Manager noted below. Do not reply to this e-mail.

Please refer to this URL address:

<http://rapidproof.cadmus.com/RapidProof/retrieval/index.jsp>

Login: Your e-mail address

Password: ----

This site contains one file. You will need to have Adobe Acrobat Reader software to read this file. This is free software that is available for user downloading at <http://www.adobe.com/products/acrobat/readstep.html>

This file contains the following:

- 1) ESA Publishing Agreement (copyright transfer form)
- 2) Reprint Order Form
- 3) Proofreading marks for your reference
- 4) Page proofs of your article and author queries, if applicable

After printing the PDF file, please read the page proofs carefully and limit any changes to those that alter scientific meaning, such as ambiguous sentences or errors in tables.

- * Mark clearly legible corrections in red directly on the page proof, preferably in the margins;
- * Answer all author queries, indicated as AQ:1, AQ:2, AQ:3, etc., in the margins of the proofs and listed on the last page of the proofs; and
- * Verify that tables, figures, and figure captions are correct and in proper sequence.

Authors are entirely responsible for marking all errors.

Any changes you make that are not the result of editor or printer error or response to a query will be billed to you at a cost of \$6.00 per affected line of text. Changes in figures will be charged at \$50.00 per figure. Color proofs of color figures are available upon request.

Publishing Agreement

Your article will not be published unless this form is completed and returned with your page proof.

Ordering Reprints

If you wish to order reprints, please use the Reprint Order Form. You can order paper reprints, a PDF reprint, or both. A PDF reprint is a PDF file of your article that is part of the online journal article and is freely accessible to anyone. You may immediately obtain the PDF file of your article from the ESA server and post it on your home page and on other publicly accessible servers. You may also use it to send your article to colleagues as an e-mail attachment and to make as many "electronic reprints" of your article as you wish. Reprint orders for articles must be received before the issue goes to press and should be mailed to the address on the form at the same time you return your page proofs.

Within 48 hours, please mail the page proofs to the address below. Also include the completed Publishing

Agreement and Reprint Order Form, otherwise publication of your article will be delayed.

Thank you for publishing with ESA. We hope that your experience has been positive. If you have any questions, contact the Environmental Entomology Issue Manager noted below.

Jane Slavin
Issue Manager
Environmental Entomology
Cadmus Professional Communications
940 Elkridge Landing Road
Linthicum, MD 21090
Phone: 410-691-6293
Fax: 410-684-2790
slavinj@cadmus.com



Environmental Entomology

Reprint Order Form – 2005

Use the form to order paper reprints, open access reprints, or both. The order form should accompany the return of your page proofs to ensure that your order is produced on time. Late orders for paper or open access reprints will be billed at much higher prices. Be sure to fill out the order form completely and sign your name where indicated under Billing Information. Reprint prices are subject to change without notice. Paper reprints are shipped within 2 weeks of the journal issue's mailing date.

1 Article Information (Please type or print clearly)

Manuscript number EN-05-6161 Number of pages _____

Reprint number **3020402**

Title _____

Author(s) _____

2 Billing Information

For all international orders, a valid credit card number is required for prepayment.

Visa Mastercard

Account No. _____ Exp. Date _____

Cardholder Signature _____

Cardholder Name _____

Cardholder Address _____

City _____ State/Province _____

Postal Code _____ Country _____

Telephone _____ Fax _____

Check/Money Order (payable to Entomological Society of America)

Purchase Order

Purchase Order must:

- accompany reprint form
- list Entomological Society of America as vendor
- include manuscript number
- include journal title, article title, and author's name

3 Reprint Prices (\$US) (Domestic shipping included)

Pages	Paper Reprints (B&W)				Open Access Reprints
	100	200	300	400	
1-2	\$ 51	56	60	66	Unlimited
3-4	87	99	107	118	38
5-8	169	187	207	228	65
9-12	243	274	306	337	126
13-16	315	356	400	444	181
Add'l 1-4 pages	79	91	102	113	236
Covers*	113	128	142	154	59
					N/A
Pages	Paper Reprints (Color)				Open Access Reprints
	185	305	426	546	
1-16	185	305	426	546	Use B&W prices
17-32	281	402	522	643	Use B&W prices

* Covers stating the title, authors, and journal issue are printed in black ink on heavy weight gray paper.

4 Reprint Information

Two kinds of reprints are offered. **B&W Paper Reprints** are printed on high-quality, uncoated paper, trimmed, and stapled at the fold. Color reprints are overruns of the printed journal pages, trimmed, and side stapled. An **Open Access Reprint** is a PDF file of your article that is posted with the online journal issue and is freely available in unlimited quantities.

Open Access Reprints \$ _____

Paper Reprints (quantity) _____ \$ _____

Covers (quantity) _____ \$ _____

Total reprint cost \$ _____

5 Shipping (International Orders Only)

\$ _____

All orders in the United States are shipped UPS, unless otherwise specified, and shipping is included in the price of the reprints.

Orders shipped outside the United States must have Expedited Shipping. Add 40% of the reprint cost for Expedited Shipping for each address. For addresses in Canada and Mexico add 20%.

Expedited Shipping provides delivery in 5-9 days and includes package tracing and proof of delivery.

6 Total Reprint Amount

\$ _____

7 Shipping Information (No PO Boxes allowed)

Ship _____ copies to:

Name _____

Address _____

City _____ State/Province _____

Postal Code _____ Country _____

Telephone _____ Fax _____

Ship _____ copies to:

Name _____

Address _____

City _____ State/Province _____

Postal Code _____ Country _____

Telephone _____ Fax _____

8 Complete and send at once with your page proofs to:

Issue Manager, Environmental Entomology
Cadmus Professional Communications
940 Elkridge Landing Road
Linthicum, MD 21090
Fax: 410-684-2790 Phone: 410-691-6293
e-mail: slavinj@cadmus.com

Proofreader's Marks

MARK	EXPLANATION	EXAMPLE
o	TAKE OUT CHARACTER INDICATED	o Your proof.
^	LEFT OUT, INSERT	u Your proof. ^
#	INSERT SPACE	# Yourproof. ^
9	TURN INVERTED LETTER	Your p ^o oof. ^
X	BROKEN LETTER	X Your p ^r oof.
vv#	EVEN SPACE	eg# A good proof.
o	CLOSE UP: NO SPACE	Your pro ^o of.
tr	TRANSPOSE	tr A ^o proof ^o good
wf	WRONG FONT	wf Your proof.
lc	LOWER CASE	lc Your ^r proof.
≡ caps	CAPITALS	Your proof. caps <u>Y</u> our proof.
ital	ITALIC	Your proof. ital <u>Your</u> proof.
rom	ROMAN, NON ITALIC	rom Your <u>proof</u> .
bf	BOLD FACE	Your proof. bf <u>Y</u> our proof.
..... stet	LET IT STAND	Your proof. stet Your proof.
out sc.	DELETE, SEE COPY	out sc. She Our proof. ^
spell out	SPELL OUT	spell out Queen (Eliz.)
#	START PARAGRAPH	# read. [Your
no #	NO PARAGRAPH: RUN IN	no # marked. → # Your proof.
L	LOWER	L [Your proof.]

MARK	EXPLANATION	EXAMPLE
⌈	RAISE	⌈ Your proof.
⌊	MOVE LEFT	⌊ Your proof.
⌋	MOVE RIGHT	⌋ Your proof.
	ALIGN TYPE	⌊ Three dogs. Two horses.
==	STRAIGHTEN LINE	= Your <u>p</u> roof.
o	INSERT PERIOD	o Your proof ^
/	INSERT COMMA	/ Your proof ^
:/	INSERT COLON	:/ Your proof ^
;/	INSERT SEMICOLON	;/ Your proof ^
∨	INSERT APOSTROPHE	∨ Your m ^o ans proof. ^
∨∨	INSERT QUOTATION MARKS	∨∨ Marked it proof ^
=/	INSERT HYPHEN	=/ A proofmark. ^
!	INSERT EXCLAMATION MARK	! Prove it ^
?	INSERT QUESTION MARK	? Is it right ^
Ⓚ	QUERY FOR AUTHOR	Ⓚ was Your proof read by ^
[/]	INSERT BRACKETS	[/] The Smith girl ^ ^
(/)	INSERT PARENTHESES	(/) Your proof 1 ^ ^
1/m	INSERT 1-EM DASH	1/m Your proof. ^
□	INDENT 1 EM	□ Your proof
▢	INDENT 2 EMS	▢ Your proof.
▣	INDENT 3 EMS	▣ Your proof.



ENTOMOLOGICAL SOCIETY OF AMERICA Publishing Agreement

Manuscript Number: EN-05-6161 Journal: Environmental Entomology

Manuscript Title: _____

Author(s): _____

1. I hereby transfer to the Entomological Society of America (ESA) full and complete ownership of any copyright I may have with respect to the work named above. If there is more than one author, I also certify that I am the duly authorized agent for any or all other authors of the work. (If the work is written as part of official duties as an employee of the United States or Canadian governments, check the box below.)
 This certifies that the work named above was prepared by an employee of the United States or Canadian governments as part of official duties and therefore cannot be copyrighted by ESA.
2. I understand that ESA assesses editorial review charges that are based upon the total number of pages published, reprint charges for any number of reprints I order, special charges for any color reproduction I require, and alteration charges for any changes deemed as author alterations made to page proofs of the article named above.

Editorial review charges are billed to the senior or corresponding author within one month of publication at rates established by the ESA Governing Board. **Current rates are listed below and are applied based on an author's membership status at the time of publication.** ESA members should ensure that their dues are up to date at the time of publication. Charges are made in full-page increments only, regardless of how much type appears on the last page of the article. Credit is given to members only when ESA is able to produce an article from a correctly formatted disk.

ESA Member Rate for articles prepared on disk:	\$48.00 per published page
ESA Members:	\$60.00 per published page
Nonmembers:	\$75.00 per published page

3. I agree to be responsible for the payment of all appropriate and relevant publication charges associated with the production of the above named article. **All publication and reprint charges are due upon receipt of invoice.** Unpaid balances more than 30 days past due are subject to a finance charge of 1.5% per month (18% per annum). Authors with unpaid balances of more than 90 days will lose their right to publish with ESA until the outstanding balance is settled. There is a \$25 fee for returned checks.
4. I have obtained written permission to use any quotations or excerpts from another work or from another's property not in the public domain or covered by fair use provisions of the U.S. Copyright law. Proper acknowledgment has been given in the article for the use of such material.

(OVER)

5. I authorize the editor to modify the manuscript as necessary to prepare the article for publication including changes in title, style, and format to conform to editorial usage, journal format, and ESA editorial style. Minor stylistic changes may be made right before publication to meet ESA editorial requirements. I understand that I may be asked to update material that appears in the article before publication.
6. I understand that I will receive page proofs of the above named article that will also show how the figures will be reproduced.
7. I agree to review and return the page proof within 48 hours of my receipt of it and to mark on the page proof any corrections that need to be made to the article. I understand that I am responsible for extra charges for any changes that are not a result of a printer or editor error or query.
8. I agree to refer to ESA all requests to republish or reprint the whole or any part of this article, including my own requests. Although ESA allows published articles to be posted on the author's web site two years after publication, ESA still holds the copyright on the original article and should be contacted for permission to republish or reprint it. ESA will respect the wishes of the author as to the use of the article or any material appearing in the article.
9. I have read the ethics in publishing statements that appear on the ESA web site (<http://www.entsoc.org/pubs/publish/ethics.html>) and affirm that this article is in compliance.
10. I understand that ESA will invoice me for the article after the issue of the journal in which the article appears is mailed and that I will make payment in full upon receipt of the invoice.
11. I hereby provide a billing address for the article mentioned above.

Name _____

Address _____

Telephone number _____ Date _____

Signature _____ Printed name _____
Author acting as agent for all other authors

A valid credit card number is required for international payments, including Canada and Mexico. Failure to do so will delay the publication of your paper.

VISA Mastercard

Card number _____ Expiration date _____

Signature _____ Printed name _____

Possible Foraging Benefits of Bimodal Daily Activity in *Proxyllocopa olivieri* (Lepeletier) (Hymenoptera: Anthophoridae)

DAPHNA GOTTLIEB,^{1,2} TAMAR KEASAR,³ AVI SHMIDA,¹ AND UZI MOTRO^{1,4}

Department of Evolution, Systematics and Ecology, Hebrew University, Jerusalem 91904, Israel

Environ. Entomol. 34(2): 000–000 (2005)

ABSTRACT We followed the daily and seasonal foraging patterns of the solitary bee *Proxyllocopa olivieri* during two springs and summers in Har Gilo, Israel. During the foraging season, the bees exhibited a clear bimodal daily activity pattern. They foraged mostly before sunrise and after sunset. We hypothesized that this activity schedule entails foraging benefits. We found that snapdragon (*Antirrhinum majus* L.) was a major source of nectar for *P. olivieri*. Nectar production rates in snapdragon were significantly higher during the morning activity period of *P. olivieri* than during mid-day. Bees of numerous other species foraged on snapdragon, but their activity periods hardly overlapped with *P. olivieri*. Foraging activity in *P. olivieri* was highest at a combination of high (25–35°C) temperatures and low (1–100 Lux) illumination levels. We suggest that *P. olivieri* benefits from foraging at dawn and dusk on snapdragon, and possibly additional food plants, because of exploitation of the large amounts of nectar produced. A nectar enrichment experiment revealed that *P. olivieri* does not cease foraging because of lack of available nectar. We suggest that the *P. olivieri*'s ability to fly at low light levels enables it to dominate a unique foraging niche.

KEY WORDS activity rhythm, foraging, interspecific competition, nectar, solitary bee

ACTIVITY RHYTHMS IN INSECTS are affected by extrinsic and intrinsic factors. Extrinsic factors are abiotic (e.g., temperature, illumination, wind) and biotic (e.g., competition, predation) variables. Intrinsic factors include physiological traits (e.g., thermoregulatory activity) and systematic affiliation (Herrera 1990).

In bees, the interplay of these factors defines five broad temporal categories of foraging activity: matinal, diurnal, afternoon, crepuscular, and nocturnal (Pittendrigh 1974). Several bee species, however, forage both during early morning and during late afternoon, thus showing a bimodal activity pattern (Gerling et al. 1983, Willmer 1988).

Several studies attribute bimodal daily activity in bees to thermal constraints. Large bees (>35–50 mg) are often able to raise their body temperature endothermically through shivering (Heinrich 1977, Stone 1994, Willmer and Stone 1997). However, when ambient temperatures are too low, they are not able to generate enough heat to enable initiation of morning flight and prolongation of evening flight (Stone and

Willmer 1989). When air temperatures are too high, bees are not able to dissipate heat generated in flight and may overheat (Chappell 1984, Willmer and Stone 1997). In environments with a wide daily range of ambient temperatures, these constraints often define a thermal window that allows activity only during morning and afternoon hours (Herrera 1990, Stone 1994, Willmer and Stone 1997). Desert and semidesert environments impose a rather narrow window of activity, because of the large temperature differences between day and night. The focus on temperature as a limiting factor to flight implies that bimodal activity may not be the bees' best foraging option, but may rather be imposed on them by their thermal physiology.

In this study, we study bimodal activity in bees as a foraging strategy and test whether it results in efficient foraging. We hypothesized that the foraging benefits of bimodal activity could include access to floral food resources that are produced bimodally (Barthell and Knops 1997, Groman and Pellmyr 1999) or that accumulate during the night and are available to early foragers (Groman and Pellmyr 1999). Such a correlation between the daily pattern of bee activity and food production was recorded for *Anthophora paupera* (Walker) foraging for pollen on *Alkanna orientalis* L. (Stone et al. 1999). Reduced competition with other foragers could provide an additional foraging benefit to bimodal activity. Such foraging benefits, together

¹ Department of Evolution, Systematics and Ecology and Center for Rationality, Hebrew University, Jerusalem 91904, Israel.

² Corresponding author: Department of Life Sciences, Ben Gurion University, POB 653 Beer Sheva 84105, Israel (e-mail: tkeasar@bgumail.bgu.ac.il).

³ Department of Life Sciences, Achva College, Mobile Post Shikmim 79800, Israel.

⁴ Department of Statistics, Achva College, Mobile Post Shikmim 79800, Israel.

with thermal constraints, may constitute a selective force favoring bimodal activity rhythms.

We tested our hypothesis using the solitary anthophorid bee, *P. olivieri*, as a model. This is a multivoltine species with palearctic distribution in desert and semidesert habitats. Newly emerged females forage for nectar only and typically mate shortly after emergence (Kronenberg 1984). After mating, they concentrate their activity on nest construction, provisioning, and oviposition. Females nest communally in slopes of clay-rich soils. They generally construct three to six cells in each nest and provide them with pollen and nectar. A single egg is laid in each cell. *P. olivieri* is well suited for our study question because it shows a clearly bimodal daily foraging pattern (Malyshev 1931, Kronenberg 1984). In addition, it is relatively large, conspicuous, and active for several months a year, facilitating observations.

Materials and Methods

Study Site. The study was carried out in Har Gilo, Israel, located southwest of Jerusalem, at an altitude of 900 m. The location is characterized by a semidesert climate, with cold winters and dry hot summers. Mean annual precipitation is 600 mm. The site has a rich fauna of pollinators (Shmida and Dukas 1990). The vegetation includes wild Mediterranean phrygana plants and cultivated plants. The main bee forage plants during the study period were *A. majus* L., *Abelia x grandiflora* (Rovelli ex Andr'), *Rosmarinus officinalis* L., *Teucrium creticum* L., and *Vitex agnus-castus* L.

Observations of Bee Activity. Field observations were carried out between April and October of 1998 and 1999. Snapdragon, a major food source for *P. olivieri* (see Figs. 2 and 3), was the main plant species observed. Forager composition at the study site was sampled at weekly intervals throughout the season. We walked along a fixed route for half an hour, four times daily, and recorded all visitors seen foraging on the flowers of snapdragon (total, 145 observations). The results of walking the strip once represent a "count." The sampling unit used here in the analyses of forager abundance is "foragers/count" (Herrera 1990).

Proxyclopa olivieri foraging activity on snapdragon was observed once a week at 15-min intervals throughout activity hours (total, 438 observations). In each 5-min observation, we recorded the number of flowers visited by a single, randomly selected, individual. In rare cases, when a bee could not be followed for 5 min (usually because it was foraging in a shady part of its food plant), we calculated its expected number of visits per 5 min based on the observed visitation rates. Observations of <30 s were discarded. We observed both males and females. We calculated "bee visitation rate," defined as the number of visits per bee per 5 min. The number of individuals observed depended on bee activity and our ability to follow single foragers. Because we usually observed only a few conspicuous foragers at a time, each individual could generally be tracked throughout the 5-min observation period. We

therefore estimated that most individuals were observed only once; hence, independence of data can be assumed. In a different set of 5-min observations, we recorded the number of flowers visited by all *P. olivieri* foraging individuals in a snapdragon patch of a known size (total, 508 observations). We used these data to calculate "flower visitation rates," defined as the number of visits per 100 flowers per 5 min. This measure is affected by the total number of foraging bees and by their rate of foraging at the time of observation.

Temperature and Illumination Recording. Temperature and light were recorded at 15-min intervals during the activity period of the bees on the same days that bees were observed. We recorded shade air temperature at the ground and light levels in the shade, 50 cm above ground. Both measurements were taken at the observation site.

Nectar Volume Sampling. Nectar was extracted from snapdragon flowers (20 per sample) using 1- or 5- μ l micropipettes of standard length (Vitrex, Modulohum I/S, Denmark). Nectar volumes were calculated from the length of the fluid column in the micropipette. Nectar was measured at biweekly intervals throughout the season. We did not record bee activity on days of nectar sampling to prevent possible effects of sampling on foraging conditions.

Both nectar production rates and nectar yields were recorded five times daily: before the beginning of *P. olivieri* morning activity, at the end of the morning activity, at mid-day, before the beginning of evening activity, and at the end of evening activity. Nectar production is the amount produced by plants in the absence of exploitation. Nectar yield (or standing crop) reflects the amount of nectar actually available to the bees and is affected by the production rate of the plant and the exploitation rate of foragers. Production rates were measured by emptying marked flowers (using a micropipette), covering them with fine-mesh gauze bags (mesh size, 1.5 mm) for a predetermined time, and measuring the accumulated nectar volume. Nectar yield was measured in uncovered flowers.

The length of the tongue of *P. olivieri* (12.3 ± 2.3 mm length from the base of the prementum to the end of glossa, $N = 4$) should allow access to any nectar in the corolla of snapdragon, and the volume recorded is assumed to approximate the volume available to foragers.

Nectar Enrichment Experiment. To test whether *P. olivieri*'s activity is limited by nectar availability, we manipulated flower patches to supply nectar for a longer time. During 20 d in August 1999, *P. olivieri* were allowed to forage in an *Abelia X grandiflora* patch, in which 1,000 flowers had been previously covered by fine-mesh gauze bags (mesh size, 1.5 mm). The covered flowers were revealed and made accessible to the bees 15 min before the end of natural morning foraging period. This created an additional nectar supply for bees in the patch. We compared the ending hour of the foraging activity between the nectar-enriched patch and an adjacent control patch without nectar manipulation. The *Abelia* patch was

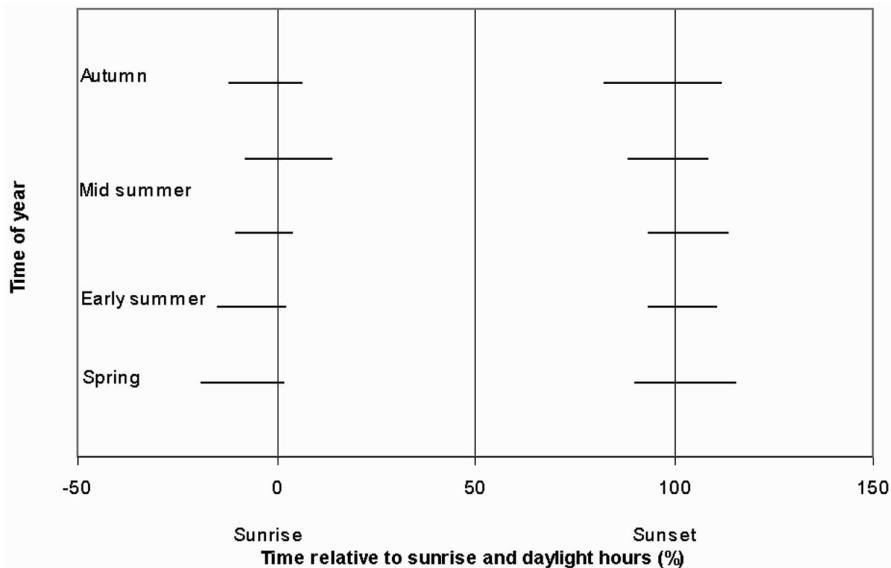


Fig. 1. Seasonal changes in the timing and duration of morning (top) and evening (bottom) foraging activity. Morning and evening activity hours are plotted relative to the times of sunrise and sunset (0 in the x-axis), respectively. Length of days (y-axis) is divided according to increasing and reducing daylength, 13–14 and 14–11 h of light, respectively. Data included only above the threshold of one visit per 5 min in 100 flowers.

visited more frequently than snapdragon at the time of the experiment and was therefore chosen as observation site.

Data Analysis. *Proxyclopa olivieri* activity was clearly correlated with daylight hours, peaking around dawn and dusk. The number of daylight hours varied between 11 and 14 during *P. olivieri*'s activity season. We corrected for this variation by presenting bee activity times relative to the time since sunrise and to the number of daylight hours on each observation day. For example, a record of activity at 0800 hours on a 14-h daylight day with sunrise at 0600 hours is reported at 14.28% of day length.

We tested the effects of temperature and light on foraging activity using two-way analysis of variance (ANOVA). We grouped light levels into three categories (1–10, 11–100, and 101–1,000 lux) and temperature records into four categories (15–20, 21–25, 26–30, and 31–35°C).

Results

Daily and Seasonal Activity Patterns. The bees showed a strongly bimodal daily foraging pattern. Foraging took place in early morning and in the evening and stopped during mid-day. Timing and duration of foraging, relative to sunrise and sunset, changed as the season progressed (Fig. 1). One apparent exception is the end of the evening activity period, which occurred between 0.75 and 1 h after sunset throughout the season (Fig. 1). In addition, duration of night nesting (according to maximal duration of activity) shortened gradually from spring (9.25 h) to autumn (7.72 h).

***Proxyclopa olivieri*'s Food Sources.** Snapdragon was a major food source for *P. olivieri* in our site,

because of its long flowering period (Fig. 2) and high attraction for the bees (Fig. 3).

Nectar production in snapdragon was significantly higher before sunrise than during daytime, with the exception of after sunset (ANOVA, $F_{(4,535)} = 4.67$; $P < 0.002$), correlating well with the morning peak in *P. olivieri* activity (Fig. 4). Nectar yields in snapdragon were not significantly affected by the time of day. The proportion of snapdragon flowers that did not contain any measurable nectar significantly increased during the day (ANOVA, $F_{(4,305)} = 3.21$, $P = 0.013$), from 18% at dawn to 38% at sunset.

Activity Period of Potential Competitors. Snapdragon was visited by numerous other foragers, mainly bees. These included honey bees (*Apis mellifera* L.), bumblebees (*Bombus terrestris* L.), carpenter bees (*Xylocopa violacea* L.), and large and small solitary bees. Body and tongue lengths for the bee species found at Har Gilo are listed in Shmida and Dukas (1990). These visitors foraged mainly during late morning and noon, with little overlap with *P. olivieri*'s activity, and exerted a high foraging pressure (Fig. 5). As a consequence, food rewards per forager visit were probably higher during *P. olivieri*'s activity periods than during the rest of the day (Fig. 4).

In the nectar enrichment manipulation, there was no significant change in the behavior of *P. olivieri*. Their activity stopped at the same time as in control patches (Fig. 6).

Abiotic Correlates of *Proxyclopa olivieri*'s Foraging Activity. High rates of foraging activity, i.e., flower visitation rates, were correlated with high air temperatures, 25–30°C ($F_{(3,365)} = 5.543$; $P < 0.001$), and low light levels, 1–100 Lux ($F_{(2,365)} = 3.708$; $P = 0.025$; Fig. 7). Mean ambient temperatures were 19.95 ± 2.99 and

F2
F3

F4

F5

F6

F7

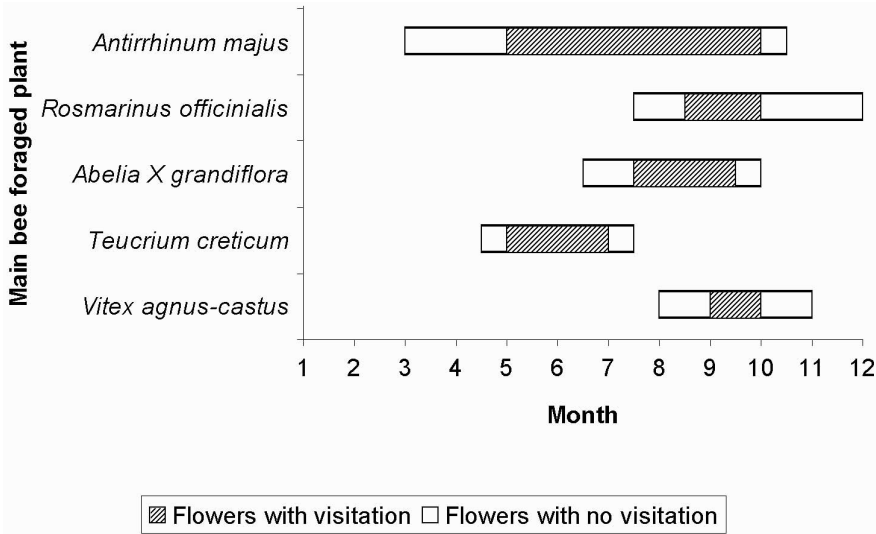


Fig. 2. Seasonal changes in flowering of the main bee forage plants and in the bees' foraging preferences. Bars indicate the flowering seasons. Periods of *P. olivieri* visitation to the flowers are indicated by the full portions of the bars. Data included only above the threshold of one visit per 5 min in 100 flowers.

$20.61 \pm 3.45^{\circ}\text{C}$ (SD) at the beginning and the end of morning activity, respectively. Mean temperatures at the beginning and the end of evening activity were, respectively, 23.82 ± 4.98 and $21.40 \pm 4.89^{\circ}\text{C}$. The significant interaction ($F_{(3,565)} = 4.09$; $P < 0.001$) between temperature and light suggests that intensity of light and temperature have a combined effect on flower visitation rates.

Discussion

The bees in our study accessed their main forage plant, snapdragon, before dawn and after dusk. Nectar production in snapdragon peaked before dawn, providing the bees with a rich food source. This nectar

production pattern might be a consequence and at the same time a cause of the early morning activity of the pollinator, as shaped by co-evolution.

Daily nectar and pollen production patterns in other food plants visited by *P. olivieri* vary widely among species (Rathcke and Real 1991, Cane 2001). It is quite possible that *P. olivieri* used more than one plant species that produces rich reward during its activity period. In addition, early morning foraging may enable *P. olivieri* to use nectar and pollen that have accumulated in flowers during the night (Groman and Pellmyr 1999). Nectar yields in snapdragon were constant throughout the day and do not seem to build up overnight. Such build-up may occur in other forage plants visited by *P. olivieri*.

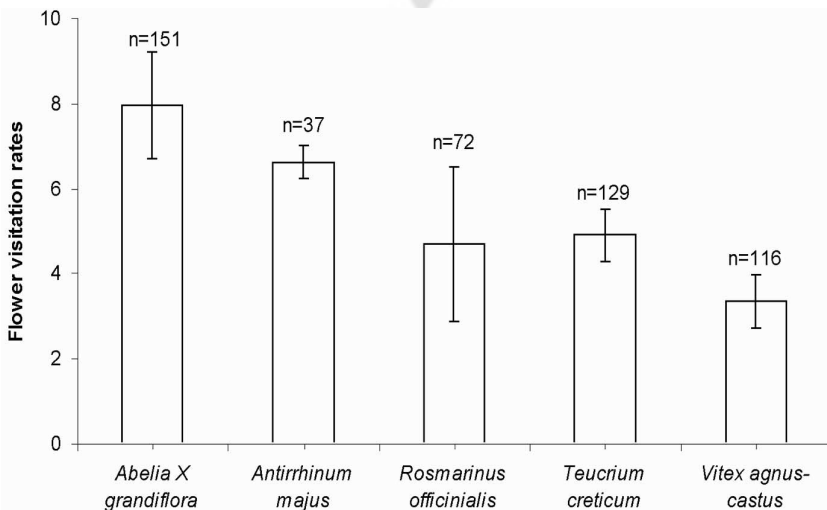


Fig. 3. Flower visitation rates at main bee foraged plants. Error bars are 1 SD.

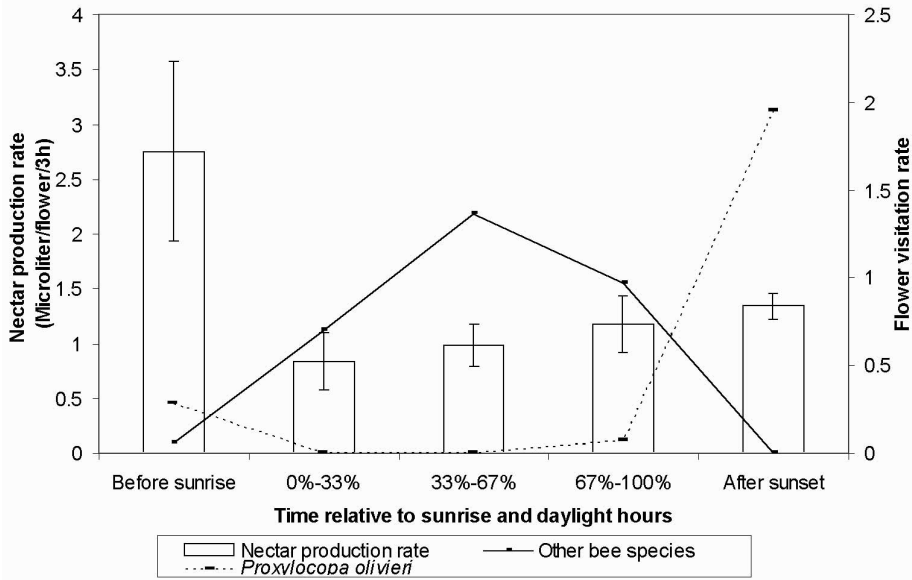


Fig. 4. Flower visitation rates and nectar production rates at different times in the day. Time plotted relative to the time since sunrise and to the number of daylight hours on each observation day. Bars: nectar production rate. Solid line: flower visitation rate for all foragers (excluding *P. olivieri*). Dotted line: flower visitation rate for *P. olivieri* (based on flower visitation rate). Error bars are 1 SD.

We also found that most of *P. olivieri*'s daily activity period did not overlap with other bee species, suggesting that their activity rhythm allowed them to evade interspecific interference competition. Interspecific competition affects the composition of bee communities and the success of invasive bee species (Buchmann 1996, Steffan-Dewenter and Tschamtko 2000, Goulson 2003). *P. olivieri*, because of its ex-

tremely bimodal activity rhythm, seems to occupy a niche that is almost competitor-free, improving its foraging prospects. The following findings imply that *P. olivieri* received a larger share of the available nectar resources by foraging early. (1) Nectar yields in snapdragon were fairly constant throughout the day, whereas nectar production was highest during *P. olivieri*'s foraging hours. The difference between

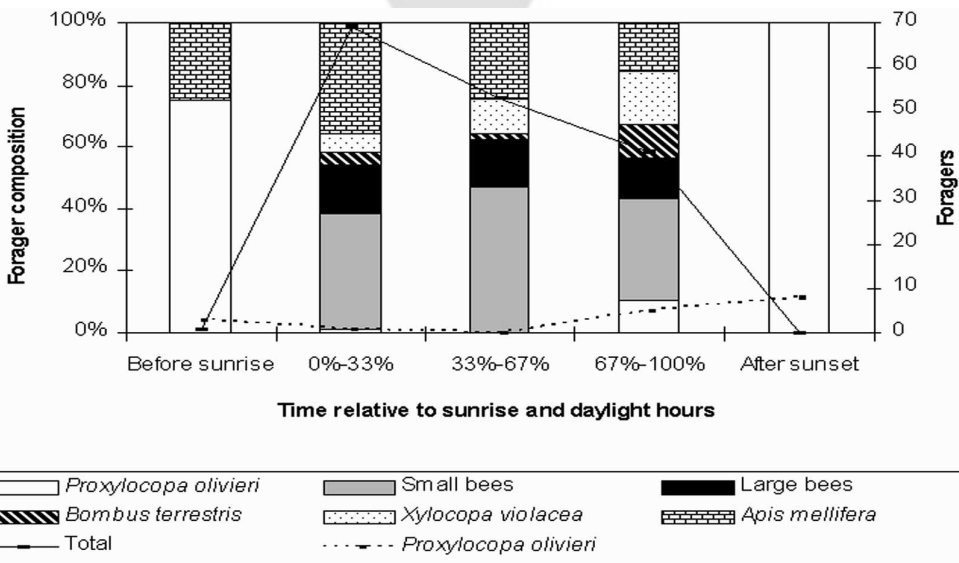


Fig. 5. Foragers/count at different times in the day, based on strip data. Time plotted relative to the time since sunrise and to the number of daylight hours on each observation day. Bars, composition of appearances by forager type; solid line, foragers/count for all foragers (excluding *P. olivieri*); dotted line, foragers/count for *P. olivieri*.

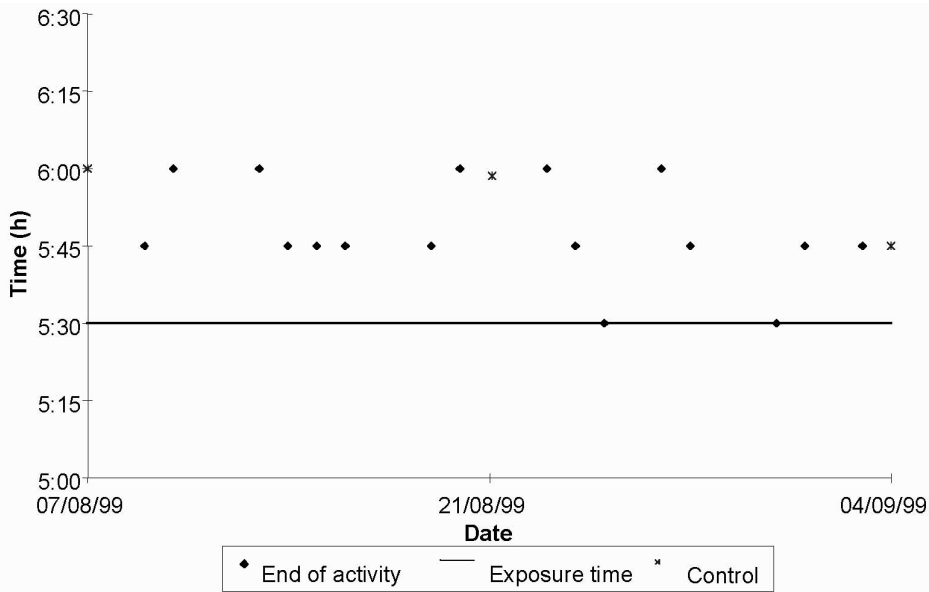


Fig. 6. End of morning foraging activity under experimental and controlled conditions. Every point represents 1 d.

production and yield, which is consumed by foragers, was therefore highest during early morning. (2) The total number of active bees was lowest during *P. olivieri*'s activity hours, allowing a higher nectar intake per foraging individual. The competitive advantage of early morning activity may be even larger for pollen foraging. This is because pollen is a depleting food resource that is likely more abundant during early morning than later in the day, whereas nectar is renewed to some extent during the day.

The working hypothesis that early morning foraging entails foraging advantages for *P. olivieri* is thus supported by high nectar production rates of its main food plant and by the low activity of its potential competitors. However, *P. olivieri*'s foraging behavior may also be constrained by its evolutionary history. For example, the mid-day heat may be a limiting factor in *P. olivieri*'s native desert habitats. Evolutionarily, bees may have been selected to be active in early morning, a pattern that persisted under the milder

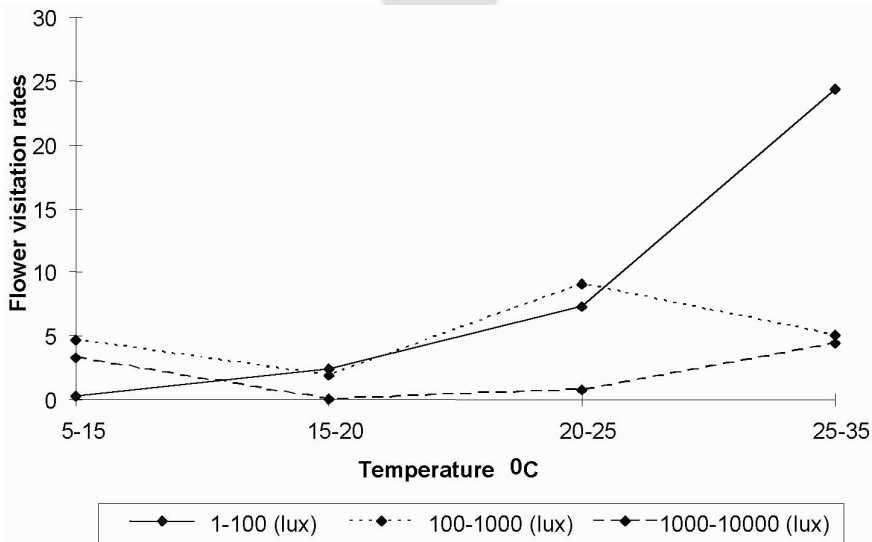


Fig. 7. Flower visitation rate at different temperatures and light intensities. Light intensities were grouped into low (1–100 Lux, bold line), medium (100–1,000 Lux, dotted line), and high (1,000–10,000 Lux, broken line). Observations <15°C were pooled into a single category because of small sample sizes.

climate conditions in this study. Such a constraint may have prevented the bees from responding to increased nectar supplies in the enrichment experiment, although temperatures were optimal. However, *P. olivieri* is active for several months a year, during which abiotic and biotic conditions change. This is expected to select for flexible foraging patterns that allow efficient performance at a wide range of environmental conditions, rather than a rigid behavioral repertoire.

Proxyclopa olivieri spends only 4–5 h/d foraging, a relatively short period compared with other bees (Dafni et al. 1987). The rest of the time is spent on nest construction and guarding. Predators and parasites are important mortality factors for broods of solitary bees (Linsley 1958, Linsley and MacSwain 1959, Stone 1994). Hence, a solitary female's nesting success is likely to be increased by a time-minimizing foraging strategy: collecting the nectar and pollen needed to provision a brood cell as quickly as possible and spending the rest of the day at the nest. Such a strategy is compatible with *P. olivieri*'s foraging pattern.

The nectar manipulation experiment that maintained artificially high levels of nectar beyond *P. olivieri*'s normal foraging hours did not change the bees' behavior. Their activity stopped at the same time as it did without the manipulation. This result supports the suggestion that *P. olivieri* does not use nectar depletion as a proximate cue to stop foraging. We assume that bees in the enriched patch were not nectar-limited, but do not know whether bees in the control patch experienced nectar shortage. Thus, one way to interpret the results is that nectar-limited and -unlimited bees have similar activity schedules. An alternative interpretation (assuming no nectar limitation in the control patch) is that foraging stopped in both patches while nectar was still abundant. In either case, we conclude that the bees used some other environmental cue (such as light intensity) to stop their morning foraging session. Although nectar depletion did not have an important proximate effect on the bees' behavior in our study, it may have been an important selective factor during the bees' evolution. In addition, it may act as a stronger proximate cue in harsher habitats, such as natural desert environments.

Foraging rates in *P. olivieri* were highest at air temperatures of 25–30°C (at low levels of illumination). Air temperatures at midday at Har Gilo are frequently within the same range, suggesting that *P. olivieri*'s inactivity at midday is not caused by bees reaching their upper thermal limit. Similarly, morning activity started, and evening activity ended, at air temperatures of 20°C, well above the lower thermal limit measured for other Mediterranean anthophorids (Stone 1994, Willmer and Stone 1997, Stone et al. 1999). We therefore suggest that ambient temperature was not a major constraint on *P. olivieri*'s activity in our study. Nevertheless, fluctuations in temperature may have caused corresponding fluctuations in nectar quality, such as nectar solute concentration (20–40 g sucrose/100 g solution), which could have affected *P. olivieri*'s behavior (Corbet 1990).

Intensity of light plays a major role not only in illumination but also in heat reflection (Corbet et al. 1993). According to this hypothesis, body temperature of bees is affected by ambient temperature and intensity of light. Direct sun is very efficient in heating up the body in comparison to heating up by ambient temperature (Stone et al. 1995). Therefore, one way of avoiding overheating of the body is to avoid direct sun (Linsley 1958). Our results agree with this suggestion, because activity increased with ambient temperature at low illumination levels only (Fig. 7). Higher light levels may bring *P. olivieri* closer to their upper thermal limit.

Foraging activity, i.e., flower visitation rates, at high temperatures was highest at very low light levels. This suggests that *P. olivieri* activity was not constrained by low levels of illumination. The ability to fly at low light levels may be better developed in *P. olivieri* than in most other bee species (Briscoe and Chittka 2001) and may allow it to occupy its unique foraging niche. However, although honey bees were not observed foraging at night at Har Gilo, they do have the ability to do so (Briscoe and Chittka 2001), hinting at a possible potential competitive interaction between *P. olivieri* and *A. mellifera*.

Exogenic factors that have been found correlating to foraging activity can act as a physiologically limiting factor to activity, but can also serve as Zeitgebers (Saunders 1982, Moore and Rankin 1993), i.e., cues that synchronize the daily cycle with external events (Saunders 1982). In this study, we have noticed two lines of evidence that may imply the existence of Zeitgebers in *P. olivieri*. (1) Evening foraging activity stopped at approximately the same time after sunset throughout the season; this suggests that sunset may serve as a Zeitgeber that couples *P. olivieri* activity pattern with a 24-h cycle. (2) Although *P. olivieri* individuals do not have continuous information on the rate of nectar production during the night, they initiate foraging before sunrise while nectar production rates are highest. To initiate foraging at the most profitable time without extrinsic information, the bee would need an inner clock.

It is possible that additional Zeitgebers can influence the timing foraging in a similar manner as light (e.g., temperature). Light/temperature manipulations are required to estimate the combined effects of these factors on the daily activity as Zeitgebers (Saunders 1982).

Acknowledgments

We thank Prof. M. Friedlander and two anonymous referees for valuable comments on the manuscript. The study was supported by the Center for Rationality and Interactive Decision Theory at the Hebrew University of Jerusalem.

References Cited

- Barthell, J. F., and J.M.H. Knops. 1997. Visitation of evening primrose by carpenter bees: evidence of a "mixed" pollination syndrome. *Southwest. Nat.* 42: 86–93.

AQ:2

AQ:3

- Briscoe, A. D., and L. Chittka. 2001. The evolution of color vision in insects. *Annu. Rev. Entomol.* 46: 471–510.
- Buchmann, S. L. 1996. Competition between honeybees and native bees in the Sonoran Desert and global bee conservation issues, pp. 125–142. *In* A. Matheson, S. L. Buchmann, C. O'Toole, P. Westrich, and I. H. Williams (eds.), *The conservation of bees*. Academic, San Diego, CA.
- Cane, J. H. 2001. Habitat fragmentation and native bees: a premature verdict? *Cons. Ecol.* 5: 3.
- Chappell, M. A. 1984. Temperature regulation and energetics of the solitary bee *Centris pallida* during foraging and intermale mate competition. *Physiol. Zool.* 57: 215–225.
- Corbet, S. A. 1990. Pollination and the weather. *Isr. J. Bot.* 39: 13–30.
- Dafni, A., D. Eisikowitch, and Y. Ivri. 1987. Nectar flow and pollinators' efficiency in two co-occurring species of *Capparis* (Capparaceae) in Israel. *Plant Syst. Evol.* 157: 181–186.
- Gerling, D., P. D. Hurd, and A. Hefetz. 1983. Comparative behavioral biology of two Middle East species of carpenter bees (*Xylocopa* Latreille) (Hymenoptera: Apoidea). *Smithson. Contrib. Zool.* 369: 1–33.
- Goulson, D. 2003. Effects of introduced bees on native ecosystems. *Annu. Rev. Ecol. Syst.* 34: 1–26.
- Groman, J. D., and O. Pellmyr. 1999. The pollination biology of *Manfreda virginica* (Agavaceae): relative contribution of diurnal and nocturnal visitors. *Oikos.* 87: 373–381.
- Heinrich, B. 1977. Why have some animals evolved to regulate a high body temperature? *Am. Nat.* 111: 623–640.
- Herrera, C. M. 1990. Daily patterns of pollinator activity, differential pollinating effectiveness, and floral resource availability in a summer-flowering Mediterranean shrub. *Oikos.* 58: 277–288.
- Kronenberg, S. 1984. Comparative exocrinology in bees with different nesting strategies: *Chalicodoma sicula*, *Proxylocopa olivieri*, *Xylocopa sulcatipes*. MSc Thesis, Tel Aviv University, Tel Aviv, Israel.
- Linsley, E. G. 1958. The ecology of solitary bees. *Hilgardia.* 27: 543–599.
- Linsley, E. G., and J. W. MacSwain. 1959. *Ethology of some Ranunculus insects with emphasis on competition for pollen*, vol. 16. University of California Publications in Entomology, University of California Press, Berkeley, CA.
- Malyshev, S. J. 1931. Lebensgeschichte der Holzbienen, *Xylocopa* Latr. (Apoidea). *Z. Morphol. Oekologie. Tiere.* 23: 754–809.
- Moore, D., and M. A. Rankin. 1993. Light and temperature entrainment of a circadian locomotor rhythm in honeybees. *Physiol. Entomol.* 18: 271–278.
- Pittendrigh, C.S. 1974. Circadian oscillations in cells and the circadian organization of multicellular systems, pp. 437–458. *In* F. O. Schmitt and F. G. Worden (eds.), *The neurosciences: third study program*. MIT Press, Cambridge, MA.
- Rathcke, B., and L. Real. 1991. Individual variation in nectar production and its effects on plant fitness in the mountain laurel (*Kalmia latifolia*). *Ecology.* 72: 149–155.
- Saunders, D. S. 1982. *Insect clocks*, 2nd ed. Pergamon, Oxford.
- Shmida, A., and R. Dukas. 1990. Progressive reduction in the mean body sizes of solitary bees active during the flowering season and its correlation with the sizes of bee flowers of the mint family (Lamiaceae), *Isr. J. Bot.* 39: 133–141.
- Steffan-Dewenter, I., and T. Tscharnkte. 2000. Resource overlap and possible competition between honeybees and wild bees in central Europe. *Oecologia (Berl.)*. 122: 288–296.
- Stone, G. N. 1994. Activity patterns of females of the solitary bee *Anthophora plumipes* in relation to temperature, nectar supplies and body size. *Ecol. Entomol.* 19: 177–189.
- Stone, G. N., and P. G. Willmer. 1989. Warm-up rates and body temperatures in bees: the importance of body size, thermal regime and phylogeny. *J. Exp. Biol.* 147: 303–328.
- Stone et al. 1995. Female foraging responses to harassment in the solitary bee *Anthophora plumipes*. *Anim. Behav.* 50: 405–412.
- Stone, G. N., F. Gilbert, P. Willmer, S. Potts, F. Semida, and S. Zalut. 1999. Windows of opportunity and the temporal structuring of foraging activity in a desert solitary bee. *Ecol. Entomol.* 24: 208–221.
- Willmer, P. G., and G. N. Stone. 1997. Temperature and water relations in desert bees. *J. Therm. Biol.* 22: 453–465.

AQ1: Please provide reference or delete citation here.

AQ2: Please provide reference or delete citation here.

AQ3: Linsley 1878 changed to 1958 to match reference list.

AQ4: Please provide first initial of first author and all authors names. et al is not allowed.



AUTHOR QUERIES

AUTHOR PLEASE ANSWER ALL QUERIES
